

Structure-function relationship of biomacromolecules: dynamics play a crucial role too

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Biomacromolecules, such as DNA, RNA, and proteins, perform complex cellular functions by adopting specific structures and/or by undergoing structural changes on relevant time scales. In this presentation, I will discuss my efforts toward understanding this structure-dynamics-function relationship of biomolecules using fluorescence spectroscopy and microscopy.

In the first part of the talk, I will present my PhD work on biophysical mechanisms of protein aggregation diseases, using amyloid- β , implicated in Alzheimer's disease, as a model peptide. I will discuss three major questions: (I) which earliest aggregate of amyloid- β is toxic?¹ (II) why is it toxic?^{2,3} and, (III) how is it toxic?^{4,5} Our results suggest that a key structural change at the earliest step of aggregation possibly drives amyloid- β toward the toxic pathway.

In the second part, I will introduce a state-of-the-art single-molecule fluorescence method called two-dimensional fluorescence lifetime correlation spectroscopy (2D FLCS).⁶ This method, developed at my postdoctoral laboratory, can reveal biomolecular structural dynamics with an unprecedented microsecond time resolution. I will talk about my contributions in further advancing this cutting-edge method, and applying it to address important biological questions. I will discuss the development of dynamic-quenching 2D FLCS, which can report microsecond-resolved local structural change of biomolecules with single dye labeling.⁷ I will also discuss the application of 2D FLCS in elucidating the folding energy landscape of the ligand-binding “aptamer” domain of prequosine riboswitch, which is a therapeutically important noncoding RNA. We propose that a microsecond structural change of the aptamer domain likely governs the biological function of the riboswitch, i.e., to regulate transcription in bacteria.⁸

References

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